

Embryogenic responses in anther derived calli of white and pink flesh guava (*Psidium guajava* L.) cultivars

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Androgenesis is reported in a few tree crops such as citrus and papaya; however, there is little information available about guava. Different modified Chu (N₆) media and cold pretreatments of floral buds were tested to induce embryogenic calli in the anthers of two white flesh guava cultivars, 'Round' and 'Pyriform'. Four modified Chu (N₆) media formulations, i.e., M₁ (400 mgL⁻¹ casein hydrolysate, 200 mgL⁻¹ glutamine, 0.2 mgL⁻¹NAA, 1.0 mgL⁻¹ KIN, 0.5 mgL⁻¹ 6-BAP, 0.5 mgL⁻¹ zeatin and 5% coconut water), M₂ (400 mgL⁻¹ casein hydrolysate, 200 mgL⁻¹ glutamine, 0.2 mgL⁻¹NAA, 1.0 mgL⁻¹ KIN, 0.5 mgL⁻¹ 6-BAP), M₁₀ (0.5 mgL⁻¹ 2,4-D), and M₁₁ (0.5 mgL⁻¹ 2,4-D and 50 mgL⁻¹ PVP) enhanced anther swelling and callus formation. Genotypic variability was found in pollen viability, phenolic exudation, and anther tissue browning after cold pretreatment for 0-72 hrs. intervals. Anthers of 'Round' showed more pollen viability across cold pretreatment intervals with less phenolic exudations and tissue browning than 'Pyriform'. On both M₁₀ and M₁₁ media, anthers with uni-nucleate microspores developed embryogenic calli; however, anthers of 'Pyriform' resulted in more calli induction (69.58%) on M₁₀ media, whereas anthers of 'Round' developed more calli (66.60%) on M₁₁ media. The proliferating calli in white flesh 'Round' and 'Pyriform' cultivars and pink flesh 'Round' cultivar formed more embryogenic masses upon transfer to growth hormone-free Chu (N₆) media under light conditions; however, embryos showed delayed maturity with limited germination. More genotypes shall be explored to enhance embryo germination, and embryogenic media shall be further fine-tuned. This study provides the foundation for the generation of homozygous lines in guava using androgenesis.

Keywords: Androgenesis, homozygosity, inbreeding, Guava, Gola, Surahi

INTRODUCTION

Guava (*Psidium guajava* L.) is a heterozygous tree fruit crop, and the diversity present in the existing seedling populations is due to the variable frequency of cross-pollination (Dinesh and Vasugi, 2016). High genetic diversity, an epigynous flower (profuse stamens of different sizes), and more juvenility limit genetic improvement in guava by conventional breeding. Moreover, selfing is laborious, time-consuming, and expensive for producing homozygous parents (Usman *et al.*, 2015; Khan *et al.*, 2017) and requires multiple selfed generations (Germanà, 2011a). Even then, it results in incomplete homozygosity (<100%), and self-incompatibility may also cause several problems (Germanà, 2006). Production of the inbred lines is the primary requirement for the development of F1 hybrids and is laborious and time-consuming in perennial fruit crops (Germana, 2011a; Niazi

and Shariatpanahi, 2020). Inbred lines may be produced more efficiently using androgenesis compared to other techniques (Heidari-Zefreh *et al.*, 2018).

In vitro, anther culture is a simple, efficient, and widely used approach for generating haploids and double haploids (Germanà, 2011a; Usman *et al.*, 2015). Androgenesis produces homozygous lines, enhances breeding efficiency, and shortens the breeding cycle. Furthermore, haploid plants serve as suitable plant material for genomic and sequencing studies (Germanà, 2011b; Zhang *et al.*, 2011a). Substantial progress has been made user *in vitro* anther culture in fruit, ornamental, medicinal, woody, and vegetable species, though response remained low in a few cases (Germanà, 2006; Wedzony *et al.*, 2009; Zhang *et al.*, 2011b; Shi *et al.*, 2012; Usman *et al.*, 2015). Doubled haploids are reported in several other fruit crops, including citrus, mulberry, papaya, apple, and custard apple (Eng and Ho, 2019).



Androgenesis has also resulted in spontaneous polyploids in fruit crops, including papaya (Germana *et al.*, 2005), apple (Hofer *et al.*, 2002), clementine (Germana 2007, 2009), and pear (Kadota and Niimi, 2004). Several factors affect the efficiency of androgenesis like donor plant's genotype, its vigor and physiological condition, culture medium compositions (Salma-Ayed *et al.*, 2010; Pencik *et al.*, 2015), combinations of plant growth regulators (PGRs), growing conditions (Bajaj, 1990; Kumar *et al.*, 2003; Qi *et al.*, 2011; Chen *et al.*, 2013), microspore development stage and pre-treatments of anthers or buds (Koleva-Guedeva *et al.*, 2007; Usman *et al.*, 2015). There is no success story of androgenesis in guava except a preliminary report about callogenesis in Indian guava genotypes (Babbar and Gupta, 1986). Hence, the current study was initiated to screen elite indigenous genotypes to establish androgenesis for the development of haploids for future breeding programs. Genotypic responses of anthers of guava cultivars to different cold shock pre-treatments, basal media composition, and PGRs are elaborated.

MATERIALS AND METHODS

A. Effect of Donor Plant, Cold Pretreatments, and Media Formulations on Androgenesis in White Flesh Guava Cultivars:

Plant Material and Media Formulations: The study was carried out during winter seasons in the Guava Germplasm Unit, Experimental Fruit Garden of the Institute of Horticultural Sciences, University of Agriculture, Faisalabad, Pakistan (N 31° 25' 46.8048", E 73° 4' 14.3112"). The average growing conditions of donor plants were 25-27°C temperature and 60%-65% relative humidity. Five healthy plants each of two white flesh guava cultivars, viz., Round 'RW' (Gola) and Pyriform 'PW' (Surahi), were selected for

androgenesis. The unopened mature and healthy floral buds (11-12 mm in length and 8-9 mm in diameter) were collected at the stage when buds swelled longer and petals appeared (Fleckinger, 1964). Most of the microspores inside the anthers are vacuolated during this phase, and it is considered the best stage for inducing gametic embryogenesis (Peixe *et al.*, 2004). Buds were exposed to 4°C temperature in the dark for inductive pre-treatment for 0, 24, 48, and 72 hours, and pollen viability was assessed. Chu (N₆) basal medium (Chu, 1978) was modified using different PGRs and vitamins (Nitsch and Nitsch, 1969) following Germana *et al.* (2011) and media pH was adjusted to 5.8 (Table 1). The media formulations were evaluated for callogenic and subsequent embryogenic responses.

Pollen Viability Assessment: Anthers were excised from cold pre-treated buds and pollens were isolated for the viability assessment. Pollen viability was determined by fine chopping of pollens in 1% acetocarmine staining dye for 3 to 5 minutes (Germana *et al.*, 2005; Abdelgadir *et al.*, 2012). Stained pollens were examined under the fluorescent microscope (CyScope Sysmex, UK) at 40X magnification and, pollen viability was calculated as given below:

$$\text{Pollen viability (\%)} = \frac{\text{No. of viable pollens counted}}{\text{Total No. of pollens}} \times 100$$

Explant Sterilization, Inoculation, and Growth Conditions: After the pollen viability test, pre-treated floral buds were sterilized by immersing them in 70% ethanol and 10% sodium hypochlorite solution containing a few drops of Tween-20 for 3 minutes, respectively, and finally rinsed in sterile distilled water thrice (Germana *et al.*, 2011; Usman *et al.*, 2015). In each petri plate, 25 anthers were inoculated on 10 ml of solid medium. Petri dishes were sealed with parafilm, incubated in the dark at 25±2 °C for the first 20 days, and then kept under fluorescent light (16/8 hrs. photoperiod).

Data was recorded for the following parameters:

Table 1. Modified Chu N₆ Medium Supplemented with Vitamins and PGRs for Androgenesis in Guava Cultivars.

Ingredients	Plant Development Media (1L)										
	M ₁	M ₂	M ₃	M ₄	M ₅	M ₆	M ₇	M ₈	M ₉	M ₁₀	M ₁₁
Chu N ₆ with Vitamins	3.99g	3.99g	3.99g	3.99g	3.99g	3.99g	3.99g	3.99g	3.99g	3.99g	3.99g
2, 4-Dichlorophenoxyacetic acid (2,4-D)	--	--	--	1mg	1mg	0.5mg	0.5mg	1mg	1mg	0.5mg	0.5mg
Polyvinylpyrrolidone (PVP)	--	--	--	--	50mg	--	50mg	--	50mg	--	50mg
Casein hydrolysate	400mg	400mg	400mg	400mg	400mg	400mg	400mg	--	--	--	--
Glutamine	200mg	200mg	200mg	200mg	200mg	200mg	200mg	--	--	--	--
Naphthaleneacetic acid (NAA)	0.2mg	0.2mg	0.2mg	0.2mg	0.2mg	0.2mg	0.2mg	--	--	--	--
Kinetin	1.0mg	1.0mg	1.0mg	1.0mg	1.0mg	1.0mg	1.0mg	--	--	--	--
6-Benzylaminopurine (6-BAP)	0.5mg	0.5mg	0.5mg	0.5mg	0.5mg	0.5mg	0.5mg	--	--	--	--
Zeatin	0.5mg	--	0.5mg	0.5mg	0.5mg	0.5mg	0.5mg	--	--	--	--
Gibberellic Acid (GA ₃)	--	--	0.5mg	0.5mg	0.5mg	0.5mg	0.5mg	--	--	--	--
Coconut Water	5%	--	5%	5%	5%	5%	5%	--	--	--	--
Sucrose	50g	50g	50g	50g	50g	50g	50g	50g	50g	50g	50g
Agar	8g	8g	8g	8g	8g	8g	8g	8g	8g	8g	8g

$$\begin{aligned} \text{Swollen yellow anthers (\%)} &= \frac{\text{No. of Yellow Swollen Anthers}}{\text{Total No. of Swollen Anthers}} \\ &\times 100 \\ \text{Swollen brown anthers (\%)} &= \frac{\text{No. of Brown Swollen Anthers}}{\text{Total No. of Swollen Anthers}} \\ &\times 100 \\ \text{Browning (\%)} &= \frac{\text{No. of Brown Anthers}}{\text{Total No. of Anthers Cultured}} \\ &\times 100 \\ \text{Callus induction (\%)} &= \frac{\text{No. of Induced Calli}}{\text{Total No. of Anthers Cultured}} \\ &\times 100 \\ \text{Embryogenesis (\%)} &= \frac{\text{No. of Germinated Embryos}}{\text{Total No. of Anthers Cultured}} \\ &\times 100 \end{aligned}$$

B. Effect of Cold Pretreatments and M₁₀ media on Androgenesis in White and Pink Flesh Guava Cultivars:

Based on the findings of the experimental study ‘A’, an extensive experiment was conducted including two white flesh (RW and PW) and one Round pink flesh (RP) guava cultivar. Anthers were collected from the promising medium-sized floral buds and pretreated for 0, 24, 48, and 72 hrs. at 4°C as discussed above. The pretreated anthers were cultured following explant sterilization conditions of experiment ‘A’ on promising media formulation M₁₀ (Table 1) for callogenesis and embryogenic responses.

Experimental Layout and Statistical Analysis: The experiment was laid under a completely randomized design (CRD) with a three-factor factorial arrangement. Differences among media, cultivars, and cold pre-treatments were tested by analysis of variance at P <0.05 level. Differences among means were tested by the Least Significant Difference (LSD) test (Steel *et al.*, 1997) for both experiments. Correlations were determined in the variables of the first experiment using R software (Gogtay and Thatte, 2017) to identify the relationship between two variables and their distribution. The inter-correlated quantitative dependent variables were analyzed by Principal Component Analysis (PCA) and Cluster Analysis (CA) (Abdi and Williams, 2010; Saraçlı *et al.*, 2013), which was conducted using Ward’s clustering method for hierarchical clustering to identify the natural clusters among variables characterized by similar attributes (Murtagh and Legendre, 2014). XLSTAT statistical software was used for multivariate analysis (Vidal *et al.*, 2020).

RESULTS

A. Effect of Donor Plant, Cold Pretreatments, and Modified N₆ Media Formulations on Anthers of White Flesh Guava Cultivars for Androgenesis:

The floral buds were stored for up to one week at 4 °C to assess the storage longevity of the anthers. The pollen of both cultivars was triangular, and genotypic variability was observed among guava cultivars for pollen viability percentage. Pollens of white flesh Round ‘RW’ cultivar showed consistently higher viability (Fig. 1C) across cold pretreatment intervals (0-72 hrs.) compared with

cv. Pyriform ‘PW’ (Fig. 2a). The pollen viability was greater (88%-94%) in freshly harvested pollens without cold treatment (0 hrs.) and viability was reduced in both Round (80%- 52%) and Pyriform (74%-44%) cultivars with the increasing interval of cold treatment from 24 to 72 hrs., respectively. The outer covering of floral buds turned blackish due to the higher phenolic contents and was slightly shrunk due to moisture loss.

After one week of culturing, both the brown (Fig. 2b) and yellow (Fig. 2c) colored anthers were swollen, and the increase in anther size continued for 5-6 weeks (Fig. 2e). Overall, browning was greater (37%-72%) in anthers cold pretreated for 0 hrs., and the browning percentage reduced with an increase in the pretreatment time intervals (24 to 72 hrs.). However, a higher percentage of yellow anthers turned brown on M₅ and M₉ media after 72-hour treatment (Fig. 2b, 3a). In contrast, the anthers maintained yellow color (no browning), and their percentage was higher in anthers treated for 24 and 48 hrs. compared with anthers treated for 0 and 72 hrs.

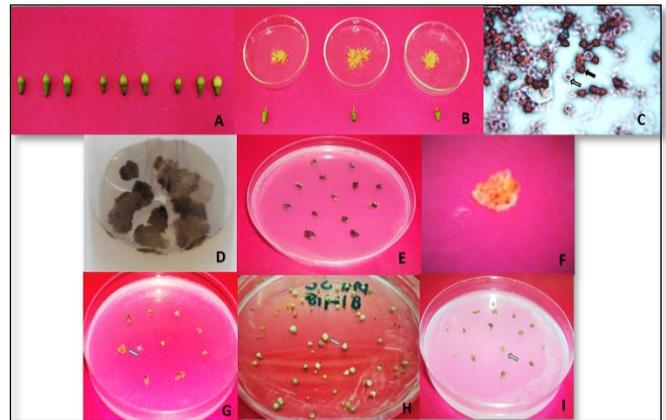


Figure 1. Androgenic responses in guava cultivars. (A) Bud selection from small, medium, and large size floral buds. (B) Anther collection from mature buds. (C) Pollen viability analysis black arrow shows viable pollens while the black lined arrow shows non-viable pollen. (D-E) Anther swelling, browning, and callus induction. (F, G) Proembryogenic calli development (grey arrows) in white flesh cv. Round anthers at M₁₀ media. (H, I) Proembryogenic calli development (grey arrows) in cv. Pyriform at M₁₁ media.

Among media treatments, the percentage of yellow anthers was maximum at M₁₀ and M₁₁ media in both Round and Pyriform cultivars after 24 hrs. of treatment, followed by M₁, M₂, M₁₀, and M₁₁ media after 48 hrs. of cold treatment (Fig. 2c, 3c). Swelling was higher in browned anthers pretreated for 0 hrs. on M₂, M₁₀, and M₁₁ media (Fig. 1D) and anthers treated for 24 hrs. on M₂ media (Fig. 2d, 3b). In contrast, the swelling was higher in yellow anthers pretreated for 24 and 48 hrs. on

M₁₀ and M₁₁ media (Fig. 2e, 3d). Overall, anther swelling percentage remained consistently higher (72%-80%) on M₁₀ and M₁₁ media at all four cold pretreatment intervals (Fig. 2f, 3e). Many swollen anthers later started callus initiation and proliferation (Fig. 1F-H).

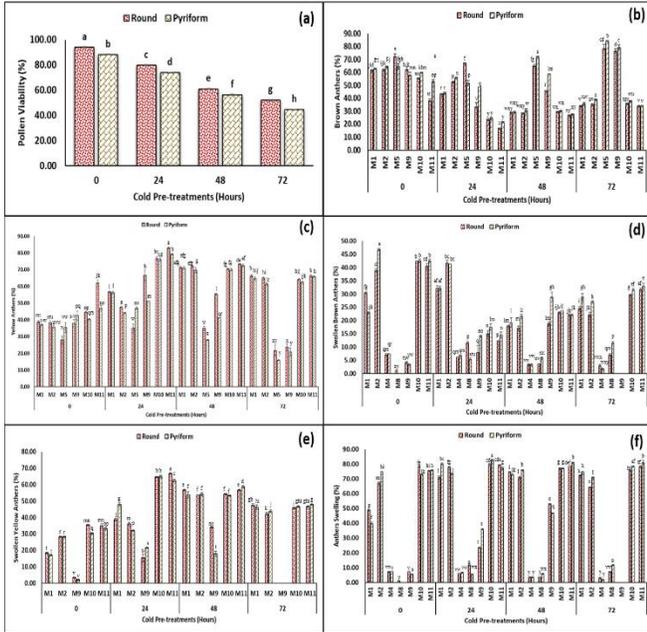


Figure 2. Effect of cold pre-treatments and modified N₆ Media on (a) Pollen viability%, (b) Brown anthers%, (c) Swollen brown anthers%, (d) Yellow anthers%, (e) swollen yellow anthers%, (f) Anthers swelling% in white flesh Guava cvs. i.e., Round and Pyriform.

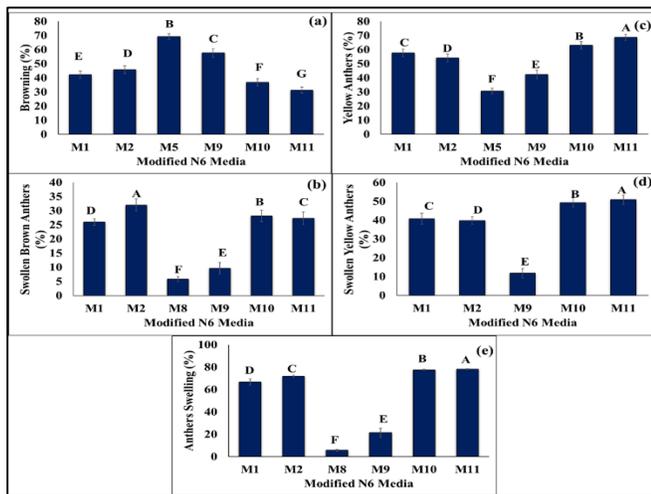


Figure 3. Overall effect of modified N₆ Media treatments on (a) Anther Browning%, (b) Swollen brown anthers%, (c) Yellow anthers%, (d) Swollen yellow anthers%, (e) Anthers swelling% in white flesh Guava cvs.

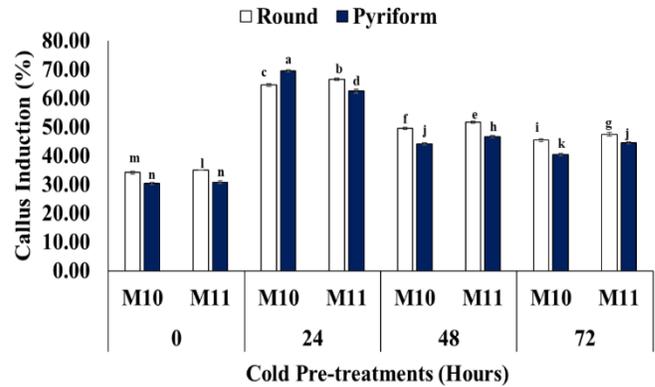


Figure 4. Embryogenic callus induction responses in cold pretreated (0, 24, 48, 72 hrs.) anthers on modified N₆ media treatments M₁₀ and M₁₁ in white flesh guava cvs. Round and Pyriform. Different alphabets show significant differences according to the LSD test at 5% probability.

Cold pretreatment of anthers for 24 hrs. significantly enhanced their potential to induce calli in both Round and Pyriform cultivars compared with control (0 hr.). The callus induction percentage was maximum in the anthers of cv. Pyriform pretreated for 24 hrs. on M₁₀ media, which decreased with an increase in the pretreatment interval from 48 to 72 hrs. (Fig. 4). Overall, among genotypes, the callus induction response was greater in cv. Round at all treatment intervals compared with cv. Pyriform on M₁₀ media. The developed calli proliferated well on the same media and grew into proembryogenic masses in anthers pretreated for 24 hrs. However, embryos showed delayed maturation and germination. Overall, anthers pre-treated at 4°C for 24 hrs. have shown a better response for callus induction on M₁₀ media. The chunks of a few developed calli turned brown after 6-8 weeks of culturing, which could be attributed to the development of more phenolic contents.

A significant positive correlation of swollen browned anthers % was observed with yellow anthers % (0.67), swollen yellow anthers % (0.73), anthers swelling % (0.89), and callus induction% (0.42). Likewise, yellow anthers % showed a significant positive correlation between swollen yellow anthers % (0.91), anthers swelling % (0.88), and callus induction% (0.61) (Fig. 5). Whereas callus induction % was positively correlated to anthers swelling % (0.63). Moreover, a negative correlation of Browning % was found between swollen brown anthers % (-0.67), yellow anthers % (-1.00), swollen yellow anthers % (-0.91), anthers swelling % (-0.88), and callus induction % (-0.61) (Fig. 5).

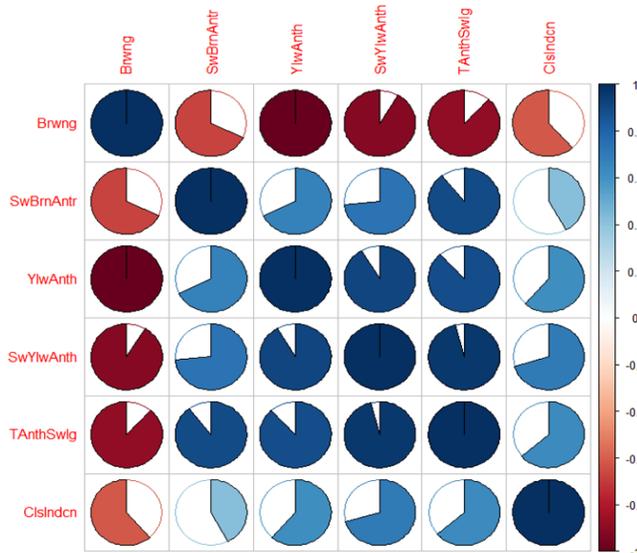


Figure 5. Correlation matrix of anther browning, swelling, callus induction responses on modified N₆ media in white flesh Guava cultivars. Negative correlations are shown in red, whereas positive correlations are shown in blue. The correlation coefficient determines the color intensity, therefore the darker the boxes are, the stronger the correlation is (i.e., the closer to -1 or 1). The correlation coefficients and the related colors are displayed in the color legend on the correlogram's right side.

PCA for callus induction rate on different media concentrations, guava cultivars, and cold pre-treatments showed great variation and was grouped into different clusters based on their characteristics. A PCA plot was developed based on first and second component factors, which showed great variation (81.55% and 10.23%) (Fig. 6a). The most divergent components and outliers in the current investigations were M₉V₁T₁, M₉V₂T₂, M₁V₂T₀, and M₂V₂T₀. A dendrogram developed based on divergence data showed the formation of two main groups (C1-C2) with nine sub-groups (A-I) and showed variation within different media concentrations, guava cultivars, and cold pre-treatments. Sub-group 'F' had a higher number of variables, while sub-groups C and I had only three variables (Fig. 6b).

B. Effect of Cold Pretreatments and Promising Culture Media (M₁₀) on Anthers of White and Pink Flesh Guava Cultivars for Androgenesis: Based on the findings discussed above, cold-pretreated anthers of three different cultivars of guava, including white flesh Round 'RW', Pyriform 'PW', and pink flesh Round (RP), were cultured on M₁₀ media for embryogenic callus induction, proliferation, and embryogenesis. The calli induced on M₁₀ media proliferated well and developed proembryogenic masses in cultivars PW and RP (Fig. 7, 8). The developed embryos showed delayed maturity and less germination, indicating a need for further tuning fine-tuning of the embryogenic medium-medium.

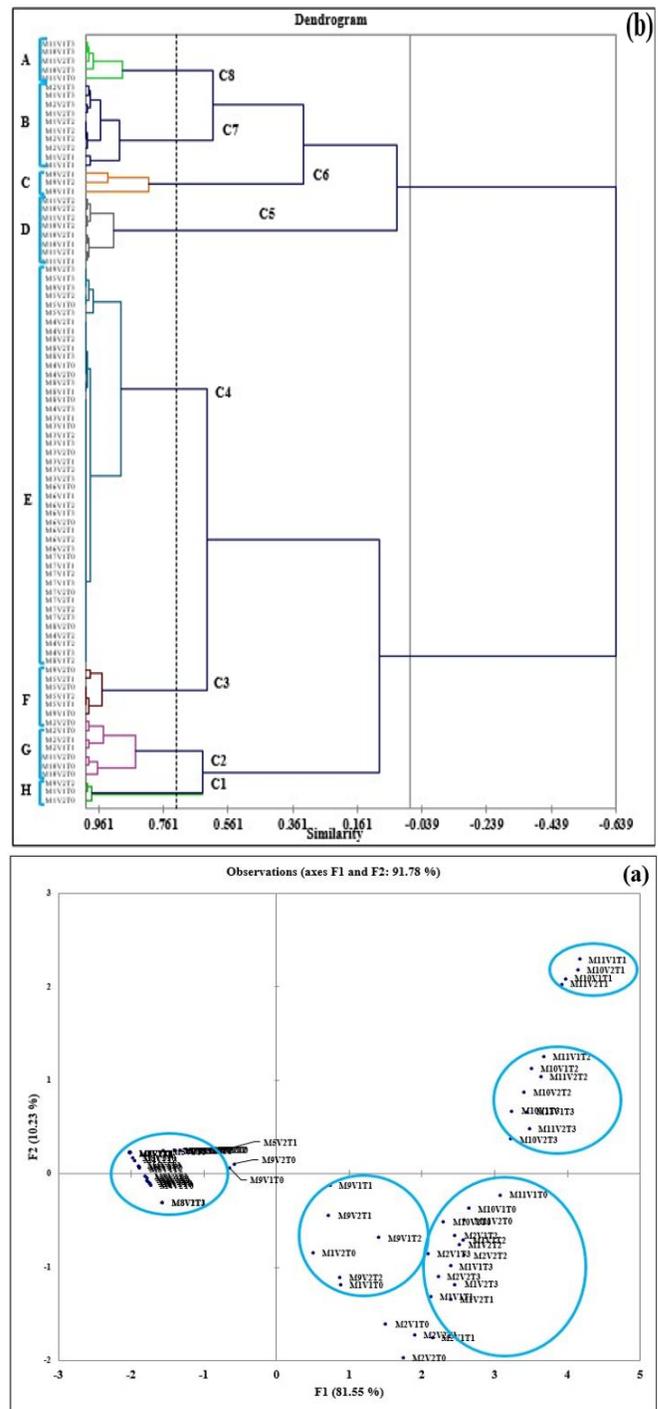


Figure 6. Principal Component Analysis (a) and Clustering Analysis (b) for different media formulations, cold pre-treatments in guava cultivars Round and Pyriform

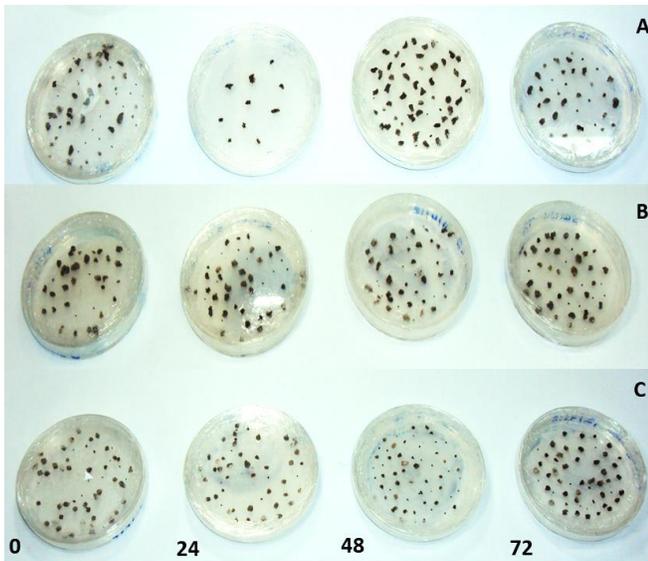


Figure 7. Proembryogenic calli development and proliferation in different guava cultivars after cold pretreatment of anthers for 0, 24, 48, and 72 hrs. on M₁₀ media (A) Round white, (B) Pyriform white, and (C) Round pink.

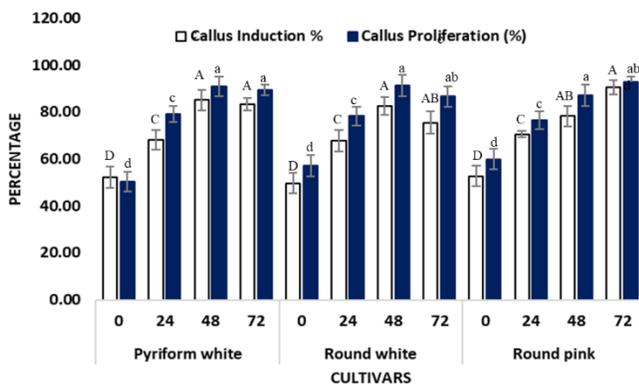


Figure 8. Embryogenic callus induction and proliferation responses in cold pretreated (0, 24, 48, 72 hrs.) anthers on modified N₆ media M₁₀ in guava cvs. Pyriform white, Round white, and Round pink.

DISCUSSION

Effect of cold treatment and genotype on pollen viability:

Pollen viability determines the success rate of the reproductive process in a specie (Abdelgadir *et al.*, 2012) and is a vital requirement for androgenesis. Androgenesis has been reported in many fruit crops, however, little is known about interaction between the water content of pollen, cold pre-treatments, and androgenic responses in guava. Staining of pollen for viability assessment is affected by plant genotype, physiology, pollen maturity, and media composition (Ferri *et al.*, 2008). Pollen grain internal

conditions like moisture content, air temperature, gametic maturity, reserve substances, and their interactions also play an important role (Pacini *et al.*, 2006). Acetocarmine has been widely used to stain pollen grains in fruit crops (Pham *et al.*, 2015; Wang *et al.*, 2015) and indicate the presence of enzymes or cytoplasm. Higher pollen variability was reported in roses (Pearson and Harney, 1984), pomegranate (Sharma and Gaur, 1984), jatropha (Lyra *et al.*, 2011), dwarf guava (Nongyai *et al.*, 2015), and litchi where 1% acetocarmine was considered more suitable for pollen viability assessment (Gupta *et al.*, 2018). In the current study, the use of 1% acetocarmine dye properly stained the pollens for viability assessment in guava cultivars and pollens of white flesh Round cultivar showed higher viability at all cold treatment time intervals. The average viability in fresh guava pollens was 91%, which is comparable to Nongyai *et al.* (2015), who reported 92% viability in dwarf ornamental guava. Pollen storage longevity varied greatly amongst cultivars, plant species, and storage conditions (Acar and Kakani, 2010; Mortazavi *et al.*, 2010). Pollen storage longevity ranged from several minutes to a few hours in cereals (Luna *et al.*, 2001) and 4 weeks in mangoes at room temperature (Dutta *et al.*, 2013). A reduction in pollen viability is also linked to moisture loss and the maintenance of a dehydrated state (Lisci *et al.*, 1994). Pollen viability in guava was reduced up to 50% after 72 hrs. pretreatment at 4°C. Pre-cold shock treatments of anthers induce callus formation, delay senescence of the anther wall, and increases the symmetric division of the pollen grains (Gueye and Ndir, 2010; Sen *et al.*, 2011). Essential substances like thermal shock proteins and amino acids for androgenesis are also released through cold treatments (Kiviharju and Pehu, 1998).

Effect of media, plant growth regulators, and sucrose on androgenic responses:

Various factors like plant genotype, pre-culture treatments, culture medium, plant growth regulators, and culture conditions (Moubayidin *et al.*, 2009; Germanà, 2011; Žur *et al.*, 2015; Usman *et al.*, 2015) can influence *in vitro* androgenesis synergistically or alone. A significant impact of cold pretreatment was observed in enhancing callus induction and reducing phenolic exudation in guava cultivars. The regeneration ability of callus depends on endogenous and exogenous concentrations of plant growth regulators (Guo *et al.*, 2017). Anthers swelling indicates the initial anatomical shift associated with the morphogenic reaction (Germanà *et al.*, 2006). Media formulations played a significant role in anther swelling, and out of eleven different media, only four media (M₁, M₂, M₁₀, M₁₁) induced anther swelling. M₁ media was additionally supplemented with zeatin and coconut water, whereas both supplements were missing in the M₂ medium. In M₁₀ and M₁₁ media formulations, all additional supplements (casein hydrolysate, glutamine, and coconut water) and PGRs (NAA, Kin, BAP, Zeatin, and GA₃) available in M₁ and M₂ media were missing (Table 1). Further, M₁₀ media was supplemented with 2,4-D,

whereas M₁₁ media had 2,4-D and PVP. The addition of 2,4-D to the anther culture medium increased the effectiveness of gametic embryogenesis in guava, as reported previously in other fruit crops (Nowaczyk *et al.*, 2016; Abdollahi and Rashidi, 2018). The lack of anther swelling in media M₃-M₉ could be attributed to the addition of GA₃ in these media, which was missing in M₁, M₂, M₁₀, and M₁₁ media inducing swelling in anthers. Similar variable callus induction responses in guava varieties were reported by Hidano *et al.* (1994) and Tsukuni (2006). Genotypic variations could also be attributed to the growing conditions of the donor plant.

Cold pre-treatments and modified Chu N₆ media affected callus initiation in tomatoes (Motallebi-Azar, 2010), citrus (Aboshama, 2011), and strawberry cultivars (Shahvali-Kohshour *et al.*, 2013). Callus induction also depends on the excised anther's nutritional requirements. Exhaustive investigations on the androgenesis using anther culture have been carried out on the type and concentration of plant growth regulators (Perera *et al.*, 2009; Smykalova' *et al.*, 2009). Regarding culture medium, Chu N₆ medium has been the most often used and effective medium for anther culture (Aboshama, 2011) and has induced gametic embryogenesis in various woody plant species like citrus, apricot, hazelnut, and loquat (Germanà and Chiancone, 2003; Karasawa *et al.*, 2016; Cardoso *et al.*, 2016). In the current study, Chu's N₆ salts and Nitsch vitamins, with or without additional supplements, PGRs, and PVP, were used for callogenesis and subsequent embryogenesis. Embryogenic callus was induced on M₁₀ and M₁₁ media having 2,4-D with or without PVP, devoid of supplements (casein hydrolysate, glutamine, and coconut water) and PGRs (NAA, Kin, BAP, Zeatin, and GA₃). In contrast, supplements of substances like casein, glutamine, coconut water, and polyvinylpyrrolidone have also been reported (Achar, 2002; Aboshama, 2011) to promote callogenesis. In fruit trees, callus induction from anthers was induced by the addition of various kinds of auxins, like NAA in *Carica papaya* (Rimberia *et al.*, 2007), and IAA in *Annona squamosa* (Nair *et al.*, 1983), 2,4-D in *Eriobotrya japonica* (Li *et al.*, 2008), 2,4-D and NAA in *Citrus clementina* (Germanà and Chiancone, 2003). Mishra and Rao (2016) stated that PGRs resulted in a high callus induction rate and reduced browning. Previously, 2,4-D has been used in anther culture medium (Abdollahi and Rashidi, 2018) and as an external pre-treatment in donor plants to increase the effectiveness of gametic embryogenesis (Nowaczyk *et al.*, 2016). Similarly, in the current study, anthers induced embryogenic calli on media (M₁₀ and M₁₁) having 2,4-D. The carbon source acts as an osmoregulatory in anther culture and has been documented in many species (Burbulis *et al.*, 2005). Elevation of sucrose concentration increased the efficiency of callus induction in different fruit crops, including citrus. Similarly, a higher level of sucrose (5%) along with suitable growth regulators increased the efficiency of callogenesis in all three guava cultivars. The embryonic initials were

observed on a few proliferating calli after 4-6 weeks of culture, whereas the rest of the calli did not develop embryogenic masses.

Conclusion: Anther culture at the uninucleate stage is found has been the most suitable stage for androgenesis. Development of embryogenic calli from anthers of guava cultivars was obtained on modified Chu N₆ media (M₁₀ and M₁₁). This study provides the baseline for establishing androgenesis in guava, whereas more genotypes and media formulations may be explored for fine-tuning the regeneration process. Furthermore, the findings of this study could lead to the development of inbred lines for breeding programs and will create new opportunities for genetic improvement of guava.

Conflict of Interest: The authors declare that there is no conflict of interest.

Author's Contribution Statements: SAMB executed the research work in the lab and prepared the article draft, BF and MU supervised the work in tissue culture cell, whereas MU, MSK, and BR conceived the idea and supervised the research plan, MU provided funds for research. MU and MSK edited the article.

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REFERENCES

- Abdelgadir, H., S. Johnson and J.V. Staden. 2012. Pollen viability, pollen germination, and pollen tube growth in the biofuel seed crop *Jatropha curcas* (Euphorbiaceae). *South African Journal of Botany* 79:132-139.
- Abdi, H. and L.J. Williams. 2010. Principal component analysis. *Wiley Interdisciplinary Reviews: Computational Statistics* 2:433-459.
- Abdollahi, M.R. and S. Rashidi. 2018. Production and conversion of haploid embryos in chickpea (*Cicer arietinum* L.) anther cultures using high 2,4-D and silver nitrate containing media. *Plant Cell, Tissue and Organ Culture* 133:39-49.
- Aboshama, H. 2011. Anther culture and plant regeneration in citrus (*Citrus volkameriana*). *Journal of Plant Production Sciences* 2:1717-1729.
- Acar, I. and V.G. Kakani. 2010. The effects of temperature on in vitro pollen germination and pollen tube growth of *Pistacia spp.* *Scientia Horticulturae* 125:569-572.
- Achar P.N. 2002. A study of factors affecting embryo yields from anther culture of cabbage. *Plant Cell, Tissue and Organ Culture* 69:183-188.

- Babbar, S.B. and S.C. Gupta. 1986. Induction of androgenesis and callus formation in *in vitro* cultured anthers of a myrtaceous fruit tree (*Psidium guajava* L.). The botanical Magazine 99:75-83.
- Bajaj, Y.P.S. 1990. Haploids in crop improvement I. Springer, Berlin.
- Burbulis, N., Blinstrubienė, A., Sliesaravičius, A. and Venskutonienė, E. 2005. Influence of genotype, growth regulators, sucrose level and preconditioning of donor plants on flax (*Linum usitatissimum* L.) anther culture. Acta Biologica Hungarica 56:323-331.
- Cardoso, J., A. Abdelgalel, B. Chiancone, R. Latado, O. Lain, R. Testolin and M. Germanà. 2016. Gametic and somatic embryogenesis through *in vitro* anther culture of different citrus genotypes. Plant Biosystems 150:304-312.
- Chen, Y., R. Ma, Y. Jiao, N. Qiao and T. Li. 2013. Impact of genotype, plant growth regulators and activated charcoal on embryogenesis induction in microspore culture of pepper. South African Journal of Botany 88:306-309.
- Chu, C.C. 1978. The N6 medium and its applications to anther culture of cereal crops. In Proceedings of Symposium on Plant Tissue Culture 43-50.
- Dinesh, M.R. and C. Vasugi. 2010. Phenotypic and genotypic variations in fruit characteristics of guava (*Psidium guajava*). Indian Journal of Agricultural Sciences 80:998-999.
- Dutta, S., M. Srivastav, R. Chaudhary, K. Lal, P. Patil, S. Singh and A. Singh. 2013. Low temperature storage of mango (*Mangifera indica* L.) pollen. Scientia Horticulturae 161:193-197.
- Eng, W.H. and W.S. Ho. 2019. Polyploidization using colchicine in horticultural plants: a review. Scientia Horticulturae 246:604-617.
- Ferri, A., E. Bellini, G. Padula and E. Giordani. 2008. Viability and *in vitro* germinability of pollen grains of olive cultivars and advanced selections obtained in Italy. Advances in Horticultural Science 22:116-122.
- Fleckinger, J. 1964. Phe´nologie et arboriculture fruitie. In: P. Grisvard and V.C. Chaudun (eds.), Le Bon Jardinier: Le Dictionnaire des plantes. Flammarion, Paris, France. pp. 362-372.
- Germana, M. and B. Chiancone. 2003. Improvement of the anther culture protocol in *Citrus clementina* hort. Ex tan. Plant Cell Reports 22:181-187.
- Germana, M., B. Chiancone, O. Lain and R. Testolin. 2005. Anther culture in *Citrus clementina*: A way to regenerate tri-haploids. Australian Journal of Agricultural Research 56:839-845.
- Germana, M.A. 2006. Doubled haploid production in fruit crops. Plant Cell, Tissue and Organ Culture 86:131-146.
- Germana, M.A. 2007. Haploidy. In: I.A. Khan (ed.), Citrus, Genetics, Breeding and Biotechnology. CAB International, Wallingford, UK. pp. 167-196.
- Germana, M.A. 2009. Haploids and doubled haploids in fruit trees. In: A. Touraev, B.P. Forster and S.M. Jain (eds.), Advances in Haploid Production in Higher Plants, Springer, Switzerland. pp. 241-263.
- Germana, M.A. 2011a. Gametic embryogenesis and haploid technology as valuable support to plant breeding. Plant Cell Reports 30:839-857.
- Germana, M.A. 2011b. Anther culture for haploid and doubled haploid production. Plant Cell, Tissue and Organ Culture 104:283-300.
- Germana, M.A., B. Chiancone, D. Padoan, I. Bárány, M.-C. Risueno and P.S. Testillano. 2011. First stages of microspore reprogramming to embryogenesis through anther culture in *Prunus armeniaca* L. Environmental and Experimental Botany 71:152-157.
- Germanà, M.A., 2016. March. Microspore embryogenesis in Citrus and other fruit crops. In IX International Symposium on In Vitro Culture and Horticultural Breeding 1187 (pp. 139-156).
- Germana, M.A., B. Chiancone, N. Levy-Guarda, P.S. Testillano and M.C. Risueno. 2006. Development of multicellular pollen of *Eriobotrya japonica* Lindl. through anther culture. Plant Science 171:718-725.
- Gogtay, N.J. and U.M. Thatte. 2017. Principles of Correlation Analysis. Journal of the Association of Physicians of India 65:78-81.
- Gueye, T. and K.N. Ndir. 2010. *In vitro* production of double haploid plants from two rice species (*Oryza sativa* L. and *Oryza glaberrima* Steudt.) for the rapid development of new breeding material. Scientific Research and Essays 5:709-713.
- Guo, B., W. He, Y. Zhao, Y. Wu, Y. Fu, J. Guo and Y. Wei. 2017. Changes in endogenous hormones and H₂O₂ burst during shoot organogenesis in TDZ-treated *Saussurea involucre* explants. Plant Cell, Tissue and Organ Culture 128:1-8.
- Gupta, A.K., M. Singh, E.S. Marboh, V. Nath and J.P. Verma. 2018. Pollen production, viability and *in vitro* pollen germination of different litchi (*Litchi chinensis*) genotypes. Indian Journal of Agricultural Sciences 88:884-888.
- Heidari-Zefreh, A.A., M.E. Shariatpanahi, A. Mousavi and S. Kalatejari. 2018. Enhancement of microspore embryogenesis induction and plantlet regeneration of sweet pepper (*Capsicum annuum* L.) using putrescine and ascorbic acid. Protoplasma 256:13-24.
- Hidano, Y., M. Niizeki and K. Saito. 1994. Production of plant by anther culture of apple. Bulletin of the Faculty of Agriculture - Hirosaki University (Japan) 58:1-21.
- Hofer, M., A. Gomez, E. Aguiriano, J.A. Manzanera and M.A. Bueno. 2002. Analysis of simple sequence repeat markers in homozygous lines in apple. Plant Breeding 121:159-162.

- Kadota, M. and Y. Niimi. 2004. Production of triploid plants of Japanese pear (*Pyrus pyrifolia* Nakai) by anther culture. *Euphytica* 138:141-147.
- Karasawa, M.M.G., B. Chiancone, V. Gianguzzi, A.M. Abdelgalel, R. Botta, C. Sartor and M.A. Germanà. 2016. Gametic embryogenesis through isolated microspore culture in *Corylus avellana* L. *Plant Cell, Tissue and Organ Culture* 124:635-647.
- Khan, H., S.C. Bhardwaj, O.P. Gangwar, P. Prasad and R. Rathore. 2017. Efficiency of double haploid production in wheat through wide hybridization and embryo rescue. *Indian Journal of Genetics and Plant Breeding* 77:428-430.
- Kiviharju, E. and E. Pehu. 1998. The effect of cold and heat pretreatments on anther culture response of *Avena sativa* and *A. Sterilis*. *Plant Cell, Tissue and Organ Culture* 54:97-104.
- Koleva-Gudeva, L.R., M. Spasenoski and T. Fidanka. 2007. Somatic embryogenesis in pepper anther culture: the effect of incubation treatments and different media. *Scientia Horticulturae* 111:114-119.
- Kumar, H.A., H. Murthy and K. Paek. 2003. Embryogenesis and plant regeneration from anther cultures of *Cucumis sativus* L. *Scientia Horticulturae* 98:213-222.
- Li, J., Y. Wang, L. Lin, L. Zhou, N. Luo, Q. Deng, J. Xian, C. Hou and Y. Qiu. 2008. Embryogenesis and plant regeneration from anther culture in loquat (*Eriobotrya japonica* L.). *Scientia Horticulturae* 115: 329-336.
- Lisci, M., C. Tanda and E. Pacini. 1994. Pollination ecophysiology of *Mercurialis annua* L. (Euphorbiaceae), an anemophilous species flowering all year round. *Annals of Botany* 74:125-135.
- Luna V.S., J. Figueroa M, B. Baltazar M, R. Gomez L, R. Townsend and J. Schoper. 2001. Maize pollen longevity and distance isolation requirements for effective pollen control. *Crop Science* 41:1551-1557.
- Lyra, D., L. Sampaio, D. Pereira, A. Silva and C. Amaral. 2011. Pollen viability and germination in *Jatropha ribifolia* and *Jatropha mollissima* (Euphorbiaceae): Species with potential for biofuel production. *African Journal of Biotechnology* 10:368-374.
- Mishra, R. and G.J.N. Rao. 2016. *In-vitro* androgenesis in rice: Advantages, constraints and future prospects. *Rice Science* 23:57-68.
- Mortazavi, S., K. Arzani and A. Moeini. 2010. Optimizing storage and *in vitro* germination of date palm (*Phoenix dactylifera*) pollen. *Journal of Agricultural Science and Technology* 12:181-189.
- Motallebi-Azar, A. 2010. Androgenic response of tomato (*Lycopersicon esculentum* Mill.) lines and their hybrids to anther culture. *Russian Agricultural Sciences* 36:250-258.
- Moubayidin, L., R. Di Mambro and S. Sabatini. 2009. Cytokinin–auxin crosstalk. *Trends in Plant Science* 14:557-562.
- Murtagh, F. and P. Legendre. 2014. Ward’s Hierarchical Agglomerative Clustering Method: Which Algorithms Implement Ward’s Criterion? *Journal of Classification* 31:274-295.
- Nair, S., P. Gupta and A. Mascarenhas. 1983. Haploid plants from *in vitro* anther culture of *Annona squamosa* Linn. *Plant Cell Reports* 2:198-200.
- Niazian, M. and M.E. Shariatpanahi. 2020. In vitro-based doubled haploid production: recent improvements. *Euphytica* 216:1-21.
- Nitsch, J.P. and C. Nitsch. 1969. Haploid plants from pollen grains. *Science* 163:85-87.
- Nongyai, N., D. Thawornchareon and K. Thaipong. 2015. Morphology, Viability and Germinability of Dwarf Guava Pollen. *Agricultural Science Journal* 46:117-120 (Original in Thailand Language).
- Nowaczyk, L., P. Nowaczyk and D. Olszewska. 2016. Treating donor plants with 2,4-dichlorophenoxyacetic acid can increase the effectiveness of induced androgenesis in capsicum spp. *Scientia Horticulturae* 205:1-6.
- Pacini, E., M. Guarnieri and M. Nepi. 2006. Pollen carbohydrates and water content during development, presentation and dispersal: a short review. *Protoplasma* 228:73-77.
- Pearson, H.M. and P.M. Harney. 1984. Pollen viability in *Rosa*. *Horticultural Science* 19:710-711.
- Peixe, A., J. Barroso, A. Potes and M.S. Pais. 2004. Induction of haploid morphogenic calluses from *in vitro* cultured anthers of *Prunus armeniaca* cv. ‘Harcot’. *Plant Cell, Tissue and Organ Culture* 77:35-41.
- Pěňčík, A., V. Turečková, S. Paulišić J. Rolčík M. Strnad and S. Mihaljević. 2015. Ammonium regulates embryogenic potential in *Cucurbita pepo* through pH-mediated changes in endogenous auxin and abscisic acid. *Plant Cell, Tissue and Organ Culture* 122:89-100.
- Perera, P.I., D. Yakandawala, V. Hoher, J.-L. Verdeil and L.K. Weerakoon. 2009. Effect of growth regulators on microspore embryogenesis in coconut anthers. *Plant Cell, Tissue and Organ Culture* 96:171-180.
- Pham, V., M. Herrero and J. Hormaza. 2015. Effect of temperature on pollen germination and pollen tube growth in longan (*Dimocarpus longan* Lour.). *Scientia Horticulturae* 197:470-475.
- Qi, Y., Y. Ye and M. Bao. 2011. Establishment of plant regeneration system from anther culture of *Tagetes patula* L. *African Journal of Biotechnology* 10:17332-17338.
- Rimberia, F.K., S. Adaniya, Y. Ishimine and T. Etoh. 2007. Morphology of papaya plants derived via anther culture. *Scientia Horticulturae* 111:213-219.

- Saraçlı, S. N. Dogan and I. Dogan. 2013. Comparison of hierarchical cluster analysis methods by cophenetic correlation. *Journal of Inequalities and Applications* 1:203.
- Sen, C., R. Singh, M. Singh and H. Singh. 2011. Effect of cold pretreatment on anther culture of boro rice hybrids. *International Journal of Plant Reproductive Biology* 3:69-73.
- Shahvali-Kohshour, R., A. Moieni and A. Baghizadeh. 2013. Positive effects of cold pretreatment, iron source, and silver nitrate on anther culture of strawberry (*Fragaria*×*Ananassa* Duch.). *Plant Biotechnology Reports* 7:481-488.
- Sharma, C.M. and R.D. Gaur. 1984. Studies on morphology, germination and viability of pomegranate (*Punica granatum* L.) pollen. *Journal of Palynology* 20:87-92.
- Shi, Q.H., P. Liu and M.J. Liu. 2012. Advances in ploidy breeding of fruit trees. *Acta Horticulturae Sinica* 39:1639-1654.
- Slama-Ayed, O., J. De Buyser, E. Picard, Y. Trifa and H.S. Amara. 2010. Effect of pre-treatment on isolated microspores culture ability in durum wheat (*Triticum turgidum* subsp. Durum Desf.). *Journal of Plant Breeding and Crop Science* 2:30-38.
- Smýkalová, I., P. Šmirous, M. Kubošiová, N. Gasmanová and M. Griga. 2009. Doubled haploid production via anther culture in annual, winter type of caraway (*Carum carvi* L.). *Acta Physiologiae Plantarum* 31:21-31.
- Steel, R.G.D., J.H. Torrie and D.A. Dicky. 1997. Principles and procedures of statistics: A biological approach. 3rd Ed. McGraw Hill Book Co., New York.
- Tsukuni, T. 2006. Production of plants derived from haploid in apple anther culture. M.Sc. thesis (unpublished). Iwate Univ., Morioka.
- Usman, M., K. Bakhsh, B. Fatima, Q. Zaman and M.H. Shah. 2015. Exploring embryogenic competence in anthers of Bitter gourd (*Momordica charantia* L.) cultivar Faisalabad long. *The Journal of Animal and Plant Sciences* 25:181-188.
- Vidal, N.P., C.F. Manful, T.H. Pham, P. Stewart, D. Keough and R. Thomas. 2020. The use of XLSTAT in conducting principal component analysis (PCA) when evaluating the relationships between sensory and quality attributes in grilled foods. *MethodsX*. 7:100835.
- Wang, L., J. Wu, J. Chen, D. Fu, C. Zhang, C. Cai and L. Ou. 2015. A simple pollen collection, dehydration, and long-term storage method for litchi (*Litchi chinensis* Sonn.). *Scientia Horticulturae* 188:78-83.
- Wedzony, M., B.P. Forster, I. Zur, E. Golemić, M. Szechynska-Hebda, E. Dubas and G. Gotebiowska. 2009. Progress in doubled haploid technology in higher plants. In: A. Touraev, B.P. Forster and S.M. Jain (eds.), *Advances in Haploid Production in Higher Plants*, Springer, Switzerland. pp. 1-34.
- Zhang, X., Q. Wu, X. Li, S. Zheng, S. Wang, L. Guo, L. Zhang and J.B.M. Custers. 2011b. Haploid plant production in *Zantedeschia aethiopica*, 'Hong Gan' using anther culture. *Scientia Horticulturae* 129:335-342.
- Zhang, S.C., A.Z. Wei and T.X. Yang. 2011a. Advances in the research and application of haploid and doubled haploid technologies in fruit trees. *International Journal of Fruit Science* 28:869-874.
- Žur, I., E. Dubas, M. Krzewska and F. Janowiak. 2015. Current insights into hormonal regulation of microspore embryogenesis. *Frontiers in Plant Science* 6:424.